Microvasculature of the harderian gland in the common

tree shrew (Tupaia glis)

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Abstract

The blood supply of the Harderian gland in common tree shrew of both sexes was studied by

using vascular corrosion cast technique in conjunction with scanning electron microscope. It was found

that the gland received the arterial blood from a branch of ophthalmic artery, which gave rise to

arterioles located in each glandular lobule, and each arteriole then further branched into capillary

plexuses that were organized into lobular appearance. Two capillary types were observed in this Harderian

gland, which are small round and large irregular sinusoidal capillaries with fenestrations. The venous

blood from this gland first drained into the orbital venous sinus locating between the glandular convex

surface and the wall of the orbit, and finally to the ophthalmic vein. The presence of the particular

vascularization together with electron dense granules and numerous stubby microvilli at the basal

portion of the acinar cells suggest that the Harderian gland in the common tree shrews may possibly

involve in endocrine function.

Key words: Harderian gland, Angioarchitecture, Vascular cast, Common tree shrew

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โครงหลอดเลือดละเอียดในต่อมฮาร์เดอร์เรี่ยนของกระแต

สมนึก นิลบุหงา, วิสุทธิ์ ประดิษฐ์อาชีพ ภาควิชากายวิภาคศาสตร์ คณะแพทยศาสตร์ มหาวิทยาลัยศรีนครินทรวิโรฒ

บทคัดย่อ

หลอดเลือดที่มาเลี้ยงต่อมฮาร์เดอร์เรี่ยน (Harderian) ของกระแต (common tree shrew) แสดง โดยใช้เทคนิค vascular corrosion cast ร่วมกับกล้องจุลทรรศน์อิเลคตรอนแบบส่องกราด (SEM) ผลการ ศึกษาพบว่าต่อมฮาร์เดอร์เรี่ยนของกระแต่ได้รับเลือดแดงมาจากหลอดเลือดออปทาลมิค (ophthalmic artery) ซึ่งแตกแขนงเป็นหลอดเลือดแดงขนาดเล็กแทรกเข้าไปในแต่ละ lobule และจะแตกแขนงย่อยต่อ เป็นหลอดเลือดแดงฝอยแบบมีรูพรุน (fenestrae) ที่ผนัง สามารถแบ่งตามลักษณะและขนาดของหลอด เลือดแดงฝอยออกเป็น 2 ลักษณะ คือ ชนิดที่เป็นท่อกลมขนาดเล็กและชนิดที่รูปร่างไม่แน่นอนขนาดใหญ่ ซึ่งหลอดเลือดแดงฝอยทั้งสองลักษณะดังกล่าวก่อรูปเป็นโครงร่างแหตามรูปร่างของหน่วยสร้างสาร ของต่อมฮาร์เดอร์เรี่ยน สำหรับเลือดดำจากต่อมฮาร์เดอร์เรี่ยนจะไหลไปยังแอ่งเลือดดำที่อยู่บริเวณตา และไหลต่อไปยังหลอดเลือดดำออปทาลมิค (ophthalmic vein) จากลักษณะที่มีหลอดเลือดแดงฝอยชนิด ที่เป็นรูพรุนจำนวนมากร่วมกับโครงสร้างของเซลล์ที่มีแกรนูลและกลุ่มของไมโครวิลไลที่ฐานล่าง จึงเป็น ไปได้ว่า ต่อมฮาร์เดอร์เรี่ยนของกระแตอาจจะทำหน้าที่เกี่ยวกับการสร้างฮอร์โมน หรือสารบางชนิด เพื่อส่งเข้าไปยังหลอดเลือดแดง

Introduction

The Harderian gland was first discovered by Jacob Harder 1 in 1694. It was described as a large structure located inferomedially of orbital cavity and was found in most terrestrial vertebrates. The gland is well developed in rodents^{2,3} for example, rat, mouse, hamster and guinea pig and also rabbit 4,5 but not in fish, bat, cow, horse, terrestrial carnivores and higher primates. 6 In adult human, the Harderian gland was absent, but not in fetus and neonate. 7-9 The secretions of mammalian Harderian gland are primarily composed of lipid, porphyrin, melatonin and inoleamine 10-13 which function in exocrine and endocrine manners. It had been proposed that the lubrication of the nictitans and cornea were considered to be the primary function of the gland. However, the actual function of this gland is still poorly understood. Even though several studies regarding Harderian gland morphology had been carried out in various species, 14-21 the detailed information concerning its blood supply is very limited. In birds, arterial blood supply of the Harderian gland comes from ophthalmotemporal branch of the external ophthalmic artery ^{22,23} whereas in rats, it comes from the inferior branch of the ophthalmic artery.²⁴ The characteristic of fenestration has been shown in the capillaries of the gland in armadillo,²⁵ mouse,²⁶ woodmouse²⁷ and golden hamster.²⁸ Its venous drainage was either via ophthalmic vein,²³ orbital venous plexus²⁹ or orbital venous sinus.^{20,30}

Three-dimensional study of the vascular pattern in the Harderian gland by the corrosion cast technique in conjunction with scanning electron microscope had not previously been performed in any animal including the common tree shrew which was considered to be a primitive primate. This study aimed to carry out two-and three-dimensional analyses of the vascular pattern of the Harderian gland in this animal.

Materials and Methods

Ten common tree shrews (Tupaia glis), five of each sex, weighing between 120-180 g were used. The preparation of the animals, the injection of Batson's #17 plastic mixture (casting medium) and the preparation of the Harderian gland vascular casts were performed according to the method previously described. 33,34 Each vascular cast of the Harderian gland was air-dried, adhered on a brass stub with silver paint, and coated with gold/palladium prior to being examined and photographed under a scanning electron microscope (SEM) at the accelerating voltage of 30 kV. For investigating by transmission electron microscopy (TEM), the Harderian glands from three females and two males were fixed by perfusion through the ascending aorta



with 2.5% glutaraldehyde in phosphate buffer, pH 7.4 and left overnight in the same fixative agent at $^{\circ}$ C. The samples were then postfixed in 1% osmium tetroxide, dehydrated in a graded series of ethanol, infiltrated and embedded in Araldite 502 resin. Semithin sections (1 μ m) were stained with toluidine blue and examined by light microscopy (LM). Thin sections (70-100 nm) were stained with 1% uranyl acetate followed by lead citrate and examined under a TEM at an accelerating voltage of 75 kV.

Results

The Harderian gland of Tupaia glis appeared to receive blood supply from a branch of ophthalmic artery (Fig. 1). The average diameter of the artery was 0.1 mm. The vascular corrosion cast technique in combination with SEM at low magnification revealed that the Harderian gland of common tree shrew was highly vascularized. The morphology of the cast conforms to the appearance of the Harderian gland (Fig. 2). When approaching the gland from the posterior end, the artery ran in the middle sulcus along the whole length of the gland (Figs. 1, 3). This sulcus was also observed on the glandular surface adjacent to eyeball and eye muscles. While running through the sulcus, the artery gave off many smaller branches, which almost ran in perpendicular to the main stem (Fig. 3). Each of these small branches, thereafter,

gave rise to 1-2 arteriole(s) to end in each glandular lobule (Fig. 4). Each arteriole further divided into plexus of true capillaries (5-7 μ m) as well as sinusoidal capillaries (10-18 μ m) (Figs. 5, 6) being organized into lobular appearance (Fig. 7). It was observed that some of the capillaries display blind ends characteristic while others continue to form the small venules (5-8 μm). These venules collected the blood into large venules (10-14 μ m) which then merged into collecting vein (17-22 µm) (Fig. 8). Approximately 3-4 collecting veins were found in each lobule and they were quite flat and ran in parallel to the surface of the gland which then emptied the blood into orbital venous sinus (Fig. 8). It was also observed that the venous blood in some venules at the glandular surface in adjacent to the orbital bones flowed directly into the orbital venous sinus (Fig. 9). This venous sinus was a broad and flat vascular space, and was seen only at the glandular surface contacting to the orbital wall (Figs. 9, 10). Moreover, the venous blood from the deep intraglandular portion drained into the collecting vein which projected directly into orbital venous sinus (Fig. 10). There were approximately 5-7 of such collecting veins in one gland.

The Harderian gland of the common tree shrew, particularly at the convex surface, was covered with the endothelial cells of the orbital venous sinus (Fig. 11).

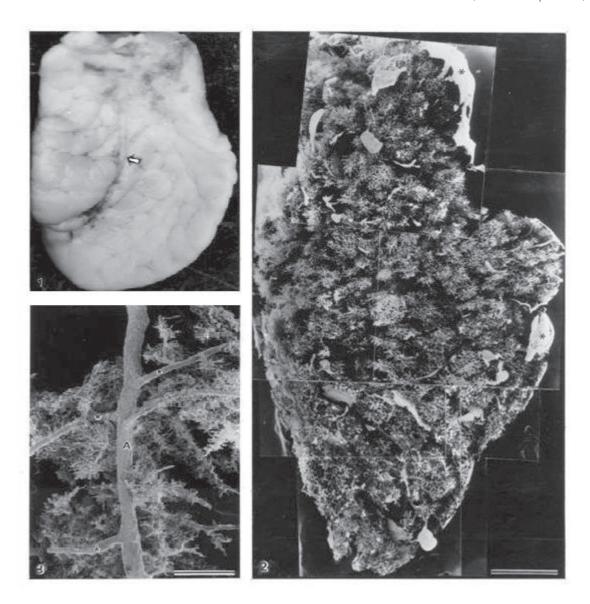


Figure 1. Picture taken from stereomicroscope of left common tree shrew Harderian gland.

This surface is adjacent to the eye sclera. Note a branch of ophthalmic artery in the middle of the gland (arrow) and the glandular lobulations. x10.

Figure 2. SEM micrograph, at low magnification, of the whole vascular cast of the common tree shrew Harderian gland at convex side. This surface is covered with orbital venous sinus (stars). The cast of orbital venous sinus, in this case, is not completely filled with plastic mixture. Bar, 1 mm.

Figure 3. SEM micrograph showing a branch of ophthalmic artery (A) running along the sulcus on the concave surface of the common tree shrew Harderian gland. The artery branches in to many small arteries (stars). Bar, 500 μ m.

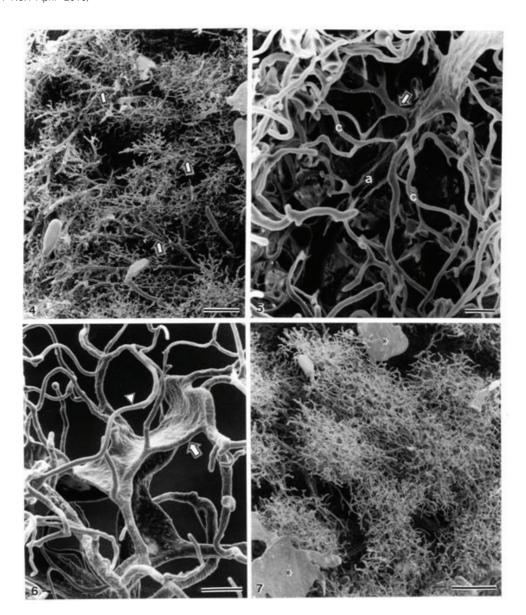


Figure 4. SEM micrograph of the vascular cast at convex surface of the common tree shrew Harderian gland after partial injection with plastic mixture, showing arterioles (arrows) branching into capillaries in each lobule. Bar, 200 μ m.

- Figure 5. SEM micrograph of an arteriole (a) branching into capillaries (c) and sinusoid (arrow) in the glandular lobule. Bar, $100~\mu m$.
- Figure 6. SEM micrograph demonstrating the casts of capillaries (arrowhead) and sinusoids (arrow) in the common tree shrew Harderian gland obtained from the razor cut specimen. Bar, $50~\mu m$.
- Figure 7. SEM micrograph showing the lobular organization of the capillaries in the common tree shrew Harderian gland at convex surface. Asterisk, part of the cast of the orbital venous sinus. Bar, 200 μ m.

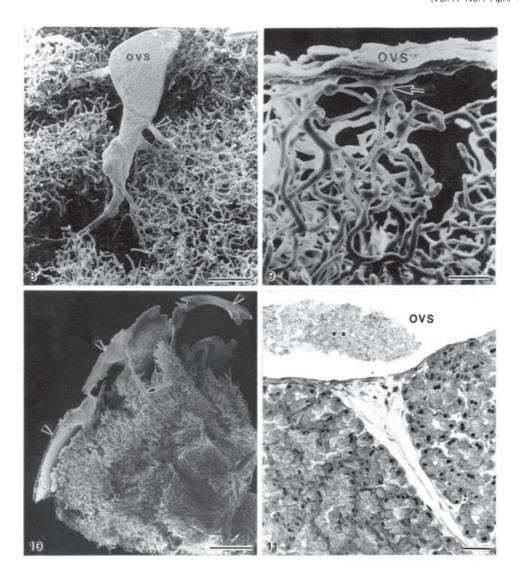


Figure 8. SEM micrograph illustrating the collecting veins (arrowheads) converge into the orbital venous sinus (OVS) at the glandular convex surface of the common tree shrew Harderian gland. Bar, 120 μ m.

Figure 9. SEM micrograph showing the venule (arrow) of some lobule drain the blood directly into the orbital venous sinus (OVS) of the common tree shrew Harderian gland. Bar, 25 μ m.

Figure 10. SEM micrograph, at low magnification, demonstrating an intraglandular vein (arrow) empties the blood into the orbital venous sinus (arrowheads) of the gland. Bar, 500 μ m.

Figure 11. Light micrograph of thick section with H&E staining showing the orbital venous sinus (OVS) filled with blood cells (stars) covering the convex surface of the Harderian gland. X10.

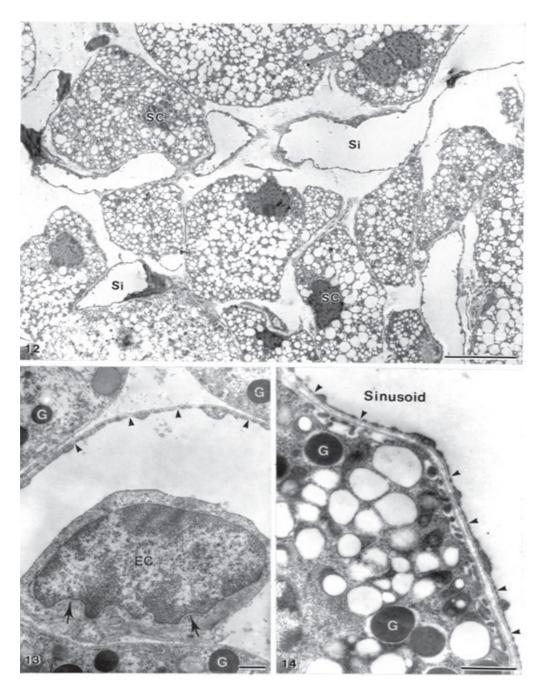


Figure 12. TEM micrograph, at low magnification, showing several fenestrated microvessels in the interstitium of the male common tree shrew Harderian gland. Note the close proximity between the sinusoids (Si) and the basal portion of secretory cells (SC). Bar, 10 μ m.

Figure 13, 14. TEM micrographs, at high magnification, of the male common tree shrew Harderian gland illustrating the capillary (Figure 13) and sinusoid (Figure 14) with fenestrations (arrowheads). Endothelial cells (EC) are usually seen with indentations (arrows) at the basal portion. Stubby microvilli at the basal portion of the secretory cell. Bar, 1 μm.

TEM micrograph showed that the gland had profused vascularization (Fig. 12). The blood vessels in the gland were of particular interest as not only small round capillaries (5-7 μ m), but also large sinusoidal capillaries (10-18 μ m) were found in various shapes (Fig. 12). These two types of capillaries were seen with fenestration (Figs. 13, 14) all over the organ. However, the population of large sinusoidal capillaries was more numerous than small round capillaries.

The capillary and sinusoidal walls were lined with an extremely flattened endothelium (Fig. 12). Frequently, the irregular sinusoidal shape adapts to the interstitial space among secretory units (Fig. 12). Those sinusoidal vessels, were located in close proximity to the acinar cells and often wedge into the basolateral portion of the cells (Fig. 12). The secretory cells contained several small electron dense granules and numerous stubby microvilli at the basolateral portion (Figs. 13, 14).

The endothelial nuclei were normally flat with minimal protrusion into the capillary lumen. The nuclear contour was relatively smooth with occasional indentations (Fig. 13). Capillary endothelial nuclei were surrounded by a small amount of cytoplasm displaying few organelles. The features of cytoplasmic expansions of the capillary endothelial cells were similar to those observed in the most endothelia. The endothelial

cell junctions could be observed and seemed to be of normal morphology. The capillary endothelium usually displayed pores intervening with thin diaphragms (Figs. 13, 14). Perforations similar to those in the hepatic sinusoids were rarely observed. It was worth noted that there was no remarkable sexual dimorphism in angioarchitecture of the common tree shrew Harderian gland.

Discussions

The Harderian gland of the common tree shew receives blood supply from a branch of ophthalmic artery similar to that of sparrow, ²³ rat ²⁴ and pig. ²⁰ In order to illustrate the three-dimensional macro and microangioarchitectures of the Harderian gland, vascular corrosion cast technique in conjunction with SEM were used in this study. With these techniques, the potassium hydroxide (KOH) is required for tissue maceration. However, as the Harderian gland tissue is composed mostly of lipid, using only KOH alone is not quite effective in tissue digestion. Therefore, a mixture of 40% KOH and 15% triton X-100 at 2:1 ratio were used instead.

Two types of capillaries were found in the Harderian gland of the common tree shrew. They were small round and sinusoidal capillaries with fenestrations. The existence of fenestrated capillaries has been previously described in the Harderian glands of nine-banded armadillo, ²⁵

mouse, 3,26,35 Sprague-Dawley rat, 21,36 woodmouse²⁷ and guinea pig.³ Sinusoidal vessels with fenestrations similar to those found in the common tree shrew have also been documented in the Harderian glands of mouse, 26 rat 36 and hamster.²⁸ However, the sinusoidal vessels in the Harderian gland of the rat, mouse and hamster appear to be less dense and smaller than those of the common tree shrew when compared with literatures. In this study, the high degree of development of vascular network and the large numbers of fenestrations of the capillaries in the interstitium of the common tree shrew Harderian gland are noteworthy. Although the precise role of sinusoids and capillaries with fenestrations found in Harderian gland of the common tree shrew are still unknown, these vessels have been generally considered as morphological evidence of an increased transport rate. 13,37 The fenestrated vessels are commonly found in endocrine organs and the liver.^{37,38} Therefore, the presence of fenestrated capillaries/ sinusoids together with the morphological features of the basal portion of the epithelial cells and the presence of stubby basal microvilli of the common tree shrew Harderian gland suggests the possibility this gland to involve in endocrine function. In other words, it suggests that there is an active transfer between the bloodstream and gland cells even though the direction of the flow

was still unknown.

The venous blood of the common tree shrew Harderian gland drains into the orbital venous sinus similar to that of mice³⁰, whereas in the case of birds²³ and rats²⁹ or piglets³⁹, it is drained via ophthalmic veins and orbital venous plexus, respectively. Because of the difference in venous drainage, drawing the blood from a mouse through the orbital venous sinus is more practical than that of a rat. The differences between orbital venous sinus and orbital venous plexus are still controversy, especially in terms of function.

References

- Harder JJ. Glandula nova lachrymalis una cum ductuexcretorio in cervis et damis. Acta Eruditorum Lipsiae 1694;49-52.
- Schreckenberger M, Reuss S. The Harderian gland of the Djungarian hamster *Phodopus* sungorus: Light and electron microscopical investigation. Acta Anat 1993;147:83-8.
- Otiz GG. Different patterns in the histology and autofluorescence of the Harderian glands of the Syrian Hamster, rat, mouse, Mongolian gerbil and guinea pig. Anat Histol Embryol 2001;30:107-15.
- 4. Ulrich H. Die venose Blutsinus in der Orbita des Kaninchens. ArchAugenheikunde 1910;65:179-88.

- Davis FA. The anatomy and histology of the eye and orbit of the rabbit. Trans Am Ophthalmol Soc Annu Meeting 1929;27: 401-41.
- 6. Payne AP. The Harderian gland: a tercentennial review. J Anat 1994;185:1-45.
- Buzzel GR. The Harderian gland: perspectives.
 Micros Res Tech 1996;34:2-5.
- Arends G, Schramm U. The structure of the human semilunar plica at different stages of its development. Amorphological and morphometric study. Ann Anat 2004;186:195-207.
- Rehorek SJ, Smith TD. The primate Harderian gland: Does it really exist? J Ann Anat 2006;188:319-27.
- Brownscheidle CM, Niewenhuis RJ.
 Ultrastructure of the Harderian gland in male
 Albino rats. Anat Rec 1978;190:735-54.
- 11. Chieffi Baccari G, Di Matteo L, Minucci S. The orbital glands of thechelonians Pseudemys scripta and Testudo graeca: Comparativehistological, histochemical and ultrastructural investigations. J Anat 1992;180:1-13.
- 12. Ranck RS, Cullen JM, Waggie KS, et al. Harderian Gland neoplasms incaptive, wild-caught beechey ground squirrels. J Vet Pathol 2008;45:388-92.

- Rubin R, Strayer DS. Rubin's Pathology. 5th
 ed. Philadelphia: Lippincott Willium & Wilkins,
 2008;618-20.
- Bodyak ND, Stepanova LV. Harderian gland ultrastructure of blacksea bottlenose dolphin (*Tursiops truncatus ponticus*). J Morphol 1994;220:207-21.
- 15. Djeridane Y. The harderian gland and its excretory Duct in the Wistar rat. A histological and ultrastructural study. J Anat 1994;184:553-6.
- 16. Coto Motes A, Rodriguez-Colunga MJ, Tolivia D, et al. Immunocytochemical localization of melatonin in the Harderian gland of Syrian hamster. Anat Rec 1996;245:13-6.
- 17. Cuthberson S, Jacson B, Toledo C, et al.
 Innervation of orbital and choroidalblood
 vessels by the pterygopalatine ganglion in
 pigeons. J Comp Neurol1997;386:422-42.
- Pradidarcheep W, Asavapongpatana S, Mingsakul T, et al. Light and electron microscopic features of the orbital Harderian gland in the common tree shrew (*Tupaia glis*).
 J Morphol 2003;255:328-36.
- Toms-Zapico C, Caballero B, Sierra V, et al. Survival mechanisms in aphysiological oxidative stress model. FASEB 2005;10.1096/ fj.04-3595fje.

- Berit HM, Hans-Jorgen S, Eldrid H, et al.
 Magnetic resonance imaging of the Harderian gland in piglets. J Anat 2006;209:699-705.
- 21. Ortiz GG, Feria-Velasco A, Pacheco-Moises FP, et al. Scanning electron microscopy of the orbital Harderian gland in the male atlantic bottlenose dolphin. Anat Histo Embryol 2009;38:279-81.
- 22. Baumel JJ. The anatomy of domestic animals, Aves heart and blood vessels 5th ed. Philadelphia: WB Saunders, 1975.
- 23. Slonaker JR. A physiological study of the anatomy of the eye and its accessory parts of the English sparrow (Passer domesticus).
 J Morphol 1981;31:351-9.
- 24. Greene EC. Anatomy of the rat. Trans Am Philos Soc 1935;27:1-370.
- 25. Weaker FJ. Light microscopic and ultrastructural features of the Harderian gland of the nine-banded armadillo. J Anat 1981;133: 49-65.
- Strum JM, Shear CR. Harderian glands in mice; fluorescence, peroxidase activity and fine structure. Tiss Cell 1982;14:135-48.
- 27. Johnston HS, McGadey J, Payne AP, et al. The Harderian gland, its secretory duct and porphyrin content in the woodmouse Apodemus sylvaticus. J Anat 1987;153: 17-30.

- 28. Menendez-Pelaez A, Toliva D, Rodriguez-Colunga MJ, et al. Ultrastructure of the blood vessels in the Harderian gland of hamster *Mesocricetus auratus:* existence of sinusoids. J Morphol 1990;204:257-63.
- 29. Timm Kl. Orbital venous anatomy of the rat.

 Lab Ani Science 1979;229:636-8.
- Cook MJ. The anatomy of the laboratory mouse. London/New york. Academic press 1965.
- Lekagul B, McNeely JA. Mammal of Thailand. Bangkok: Kurusapa Ladprao Press 1977;1-7.
- 32. Palley LS, Schlossman SF, Letoin NL.

 Common tree shrews and primate share
 leukocyte membrane antigens. J Med
 Primatol 1984;13:67-71.
- 33. Chunhabundit P, Somana R. Scanning electron microscopy on pineal vascularization of the common tree shrew (*Tupaia glis*). *J* Pineal Res 1991;10:59-64.
- 34. Promwikorn W, Thongpila S, Pradidarcheep W, et al. Angioarchitecture of the celiac sympathetic ganglion complex in the common tree shrew (*Tupaia glis*). *J* Anat 1998;193:409-16.
- 35. Carriere C. Ultrastructural visualization of intracellular porphyrin in the rat Harderian gland. Anat Rec 1985;213:496-504.

- 36. Sakai T. Major ocular glands (Harderian gland and lacrimal gland) of the must shrew *Suncus minus* with a review on the comparative anatomy and histology of the mammalian lacrimal gland. J Morphol 1989;201:39-57.
- 37. Fawcett DW. Textbook of histology 2nded.
 Philadephia: W.B. Saunders,1981.
- 38. Rattanachaikunsopon P, Chunhabundit P, Bamroonwong S, et al. Microvasculature of the thyroid gland in the common tree shrew (Tupaia glis): Microvascular corrosion cast/SEM. Acta Anat 1991;142:208-14.
- 39. Munkeby BH, Smith HJ, Winther-Larssen EH, et al. Magnetic resonance imaging of the Harderian gland in piglets. J Anat 2006;209:699-705.